



Multiple semi-continuous chemical detection by bacterial bioreporters in a microfluidics chemostat

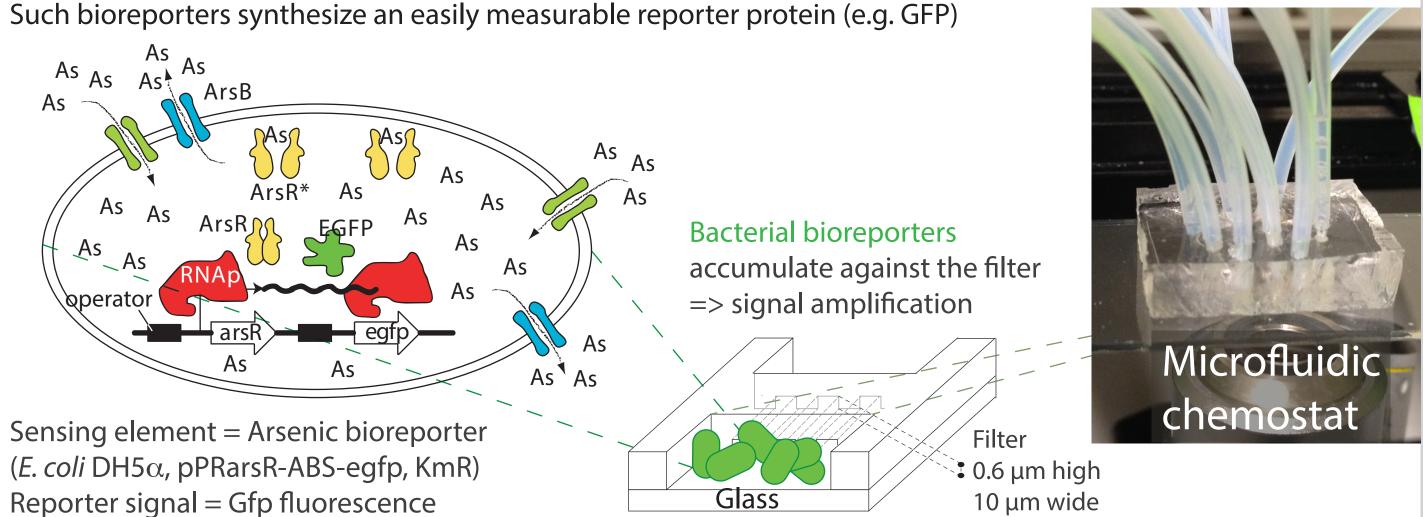
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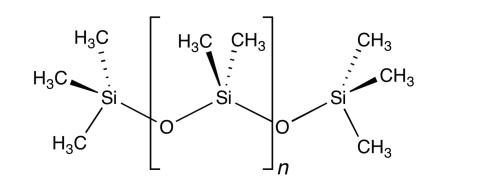
Autonomous cell-based sensing microsystem

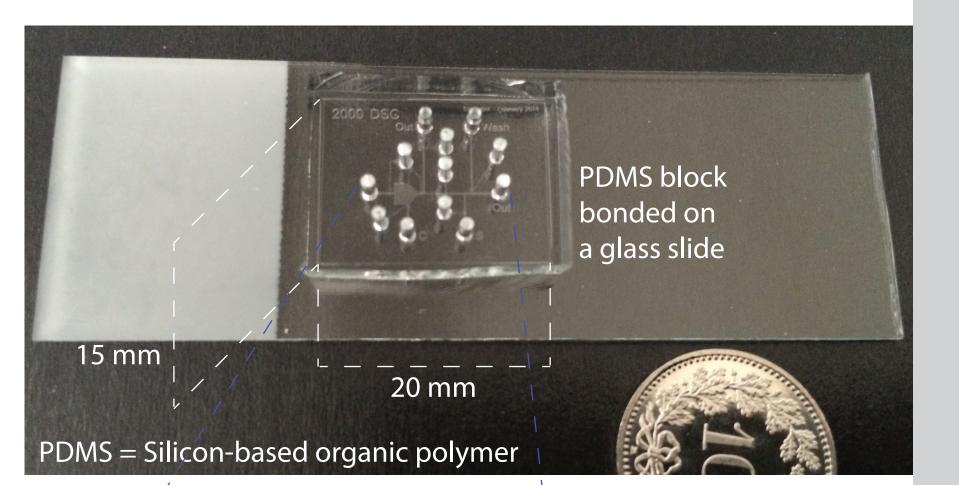
Whole-cell bacterial bioreporters are genetically-modified for the detection of different groups of chemicals (e.g. arsenite)

Such bioreporters synthesize an easily measurable reporter protein (e.g. GFP)

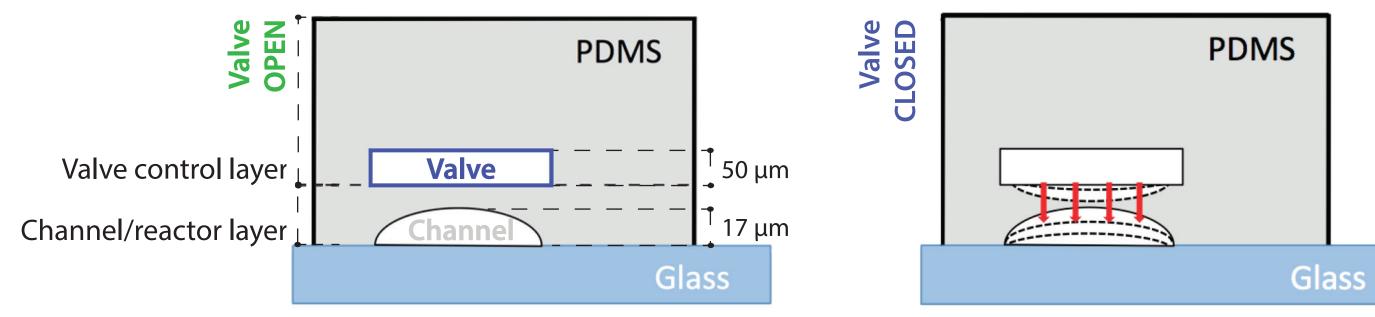


Two-layered PDMS block





Side view - valve actuation principle



Two layer PDMS chip cross section showing the valve location with the upper control channel and the lower flow channel

As a pressure is applied in the upper channel, the thin membrane deflects and close the lower channel

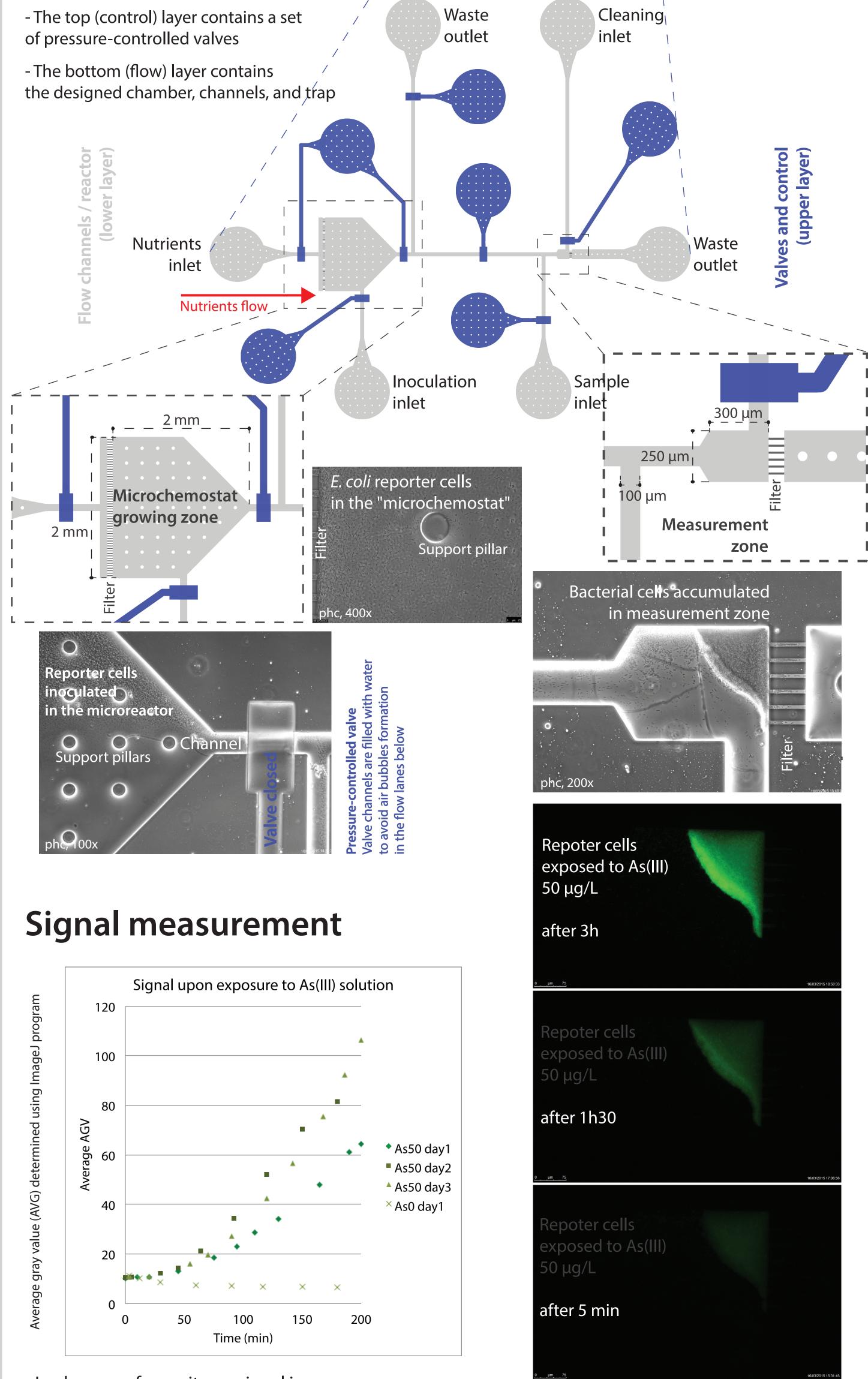
- Solenoid valves allow switching air pressure and driving the opening or closing of the PDMS valves; valves can be closed by applying air-pressure of 1.5 bar

- The different flows in the bottom layer are also driven by air-pressure

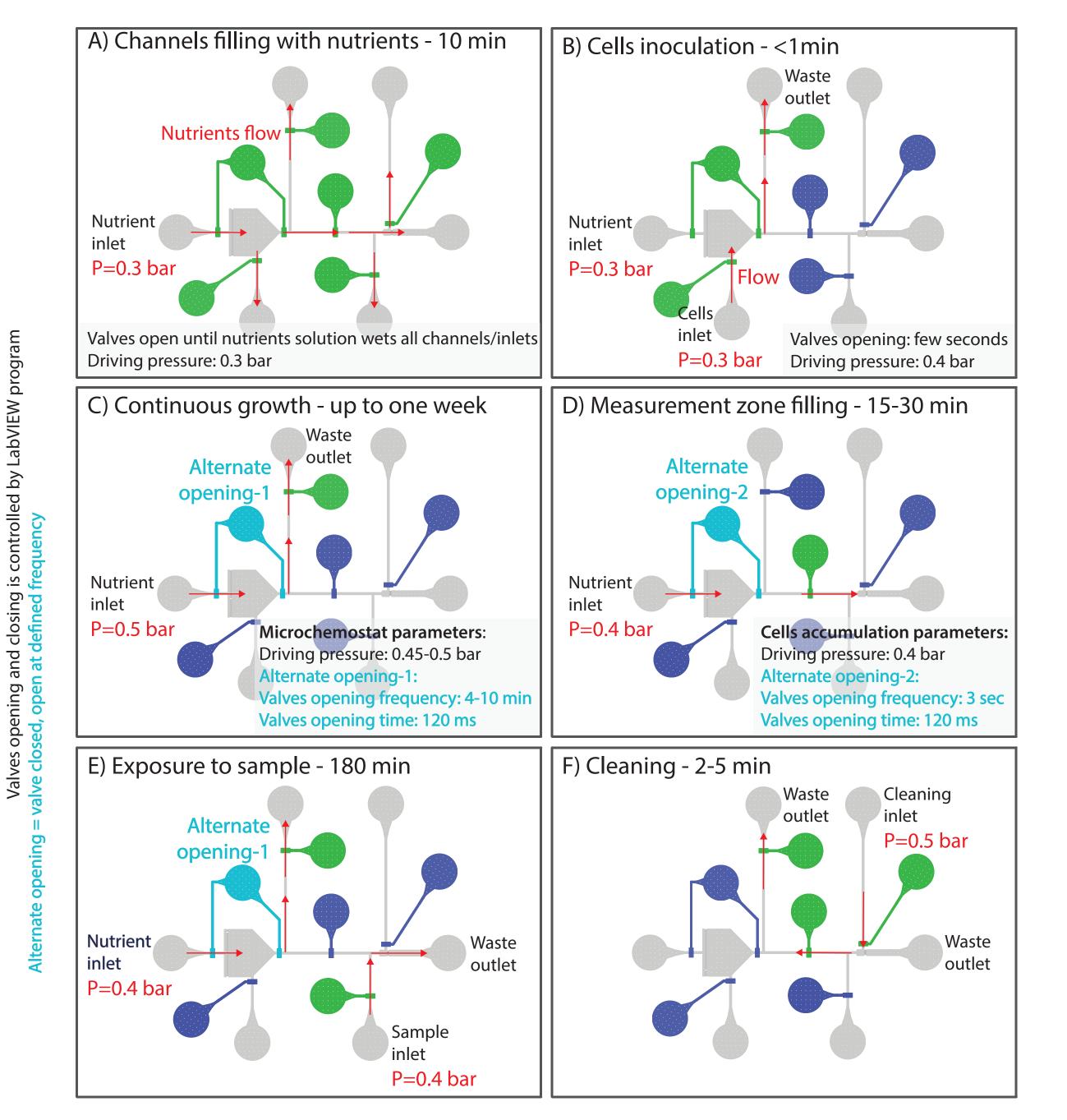
Polydimethylsiloxane (PDMS)

- Useful rheological (or flow) properties - Easy to produce
- Permeable to oxygen
- Optically clear (transparent)
- Inert, non-toxic

Top view - schematic layout (Mask/PDMS mold)



Operating the microfluidic system



C) Maintaining cells actively growing imply discontinuous addition of fresh nutrients and removal of extra cells/medium

E) Continuous growth while cells exposed to the sample during 180 min

F) Measurement zone and channels cleaning before and after each cells exosure to the sample

- In absence of arsenite no signal increase

- The out put signal of cells exposed to As(III) at 50 μ g/L is comparable over days

Conclusions

- Bioreporter cells can be maintained actively growing for one week - Multiple exposures to contaminated samples is possible - Signal can be measured by an opto-electronic device (Hes-so Valais) Acknowledgment: control and flow wafers fabrication as well as PDMS chip fabrication (soft lithography) were performed at CMi, EPFL





